



Synthesis and Biological Evaluation of an Electronically Activated Isooxacephem

Gholam H. Hakimelahi,^{a,*} Shwu-Chen Tsay,^a Hsi-Hwa Tso,^a Zahra Ramezani^b
and Jih Ru Hwu^{a,c,*}

^aOrganosilicon and Synthesis Laboratory, Institute of Chemistry, Academia Sinica, Nankang, Taipei, Taiwan 11529, R.O.C.

^bSt Thomas Research Training Laboratory, Dominican International School, Tah Chih, Taipei, Taiwan 104, R.O.C.

^cDepartment of Chemistry, National Tsing Hua University, Hsinchu, Taiwan 30043, R.O.C.

Abstract—New isooxacephem (\pm)-3-ethyl 2-hydrogen (6*RS*,7*RS*)-8-oxo-7-(phenylacetamido)-4-oxa-1-azabicyclo[4.2.0]oct-2-ene-2,3-dicarboxylate (**8**) was synthesized from (\pm)-dibenzyl 2-[*cis*-2-oxo-3-(phenylacetamido)-4-styryl-1-azetidiny]-2-[*t*-butyldimethylsiloxy(methoxycarbonyl)methyl]malonate (**1**) in six steps. This bicyclic β -lactam was found to possess notable biological activities against several pathogenic microorganisms in vitro, including *Staphylococcus aureus* 95, *S. aureus* FDA 209P, *Escherichia coli* ATCC 39188, *Salmonella typhi* O-901, *Pseudomonas aeruginosa* 18S-H, *P. aeruginosa* 1101-75, and *Klebsiella pneumoniae* NCTC 418. The electronic activation of the β -lactam moiety by an ester group plays a prominent role in the biological activity of this novel isooxacephem. Copyright © 1996 Elsevier Science Ltd

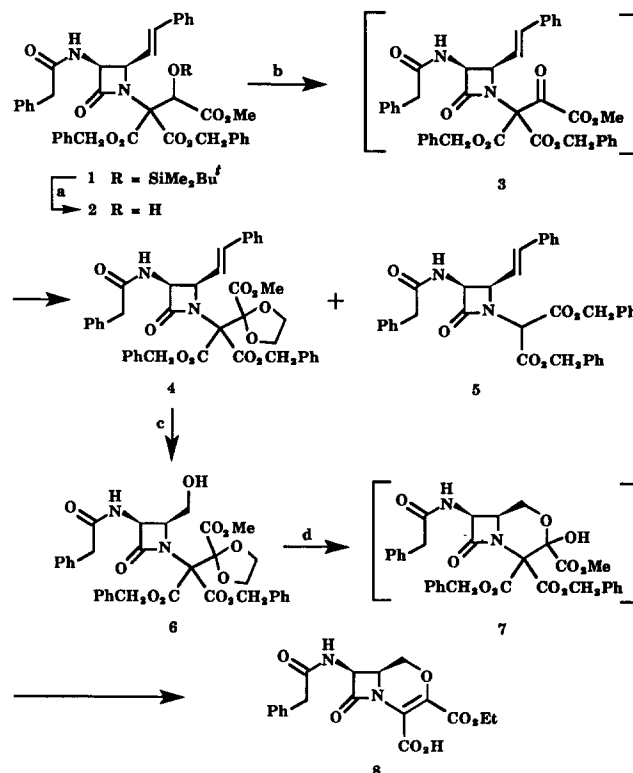
Introduction

Many β -lactams with a fused heterocyclic B-ring have been used in medicine and pharmacology.¹ Continuous efforts are needed in the development of new strategies for synthesis of novel β -lactams and in the exploration of structural modification for enhancement of their biological activities.

We synthesized a new isooxacephem (**8**) by utilizing the strategy involving construction of the 3,4-dihydro-2*H*-[1,4]oxazine ring (Scheme 1). We found that this isooxacephem possessed appealing antibacterial activity.

Results and Discussion

We desilylated silyloxy β -lactam (\pm)-**1**² with Bu₄NF in THF to give the corresponding alcohol **2** in 95% yield. Oxidation of **2** by use of 4-methylmorpholine *N*-oxide in the presence of a catalytic amount of tetrapropylammonium perruthenate³ in CH₂Cl₂ afforded ketoester **3**. Upon treatment with a benzene solution of ethylene glycol and a catalytic amount of *p*-toluenesulfonic acid in situ, **3** was led to a mixture including the desired acetal **4** (23% overall yield) and the decarbonylated product **5**⁴ (60% overall yield). Ozonolysis of **4** with an oxygen–ozone mixture in MeOH followed by reduction with NaBH₄ gave alcohol **6** in 10% yield only. We improved the yield to 46% by introducing ozone and nitrogen gases simultaneously;⁵ the reaction mixture

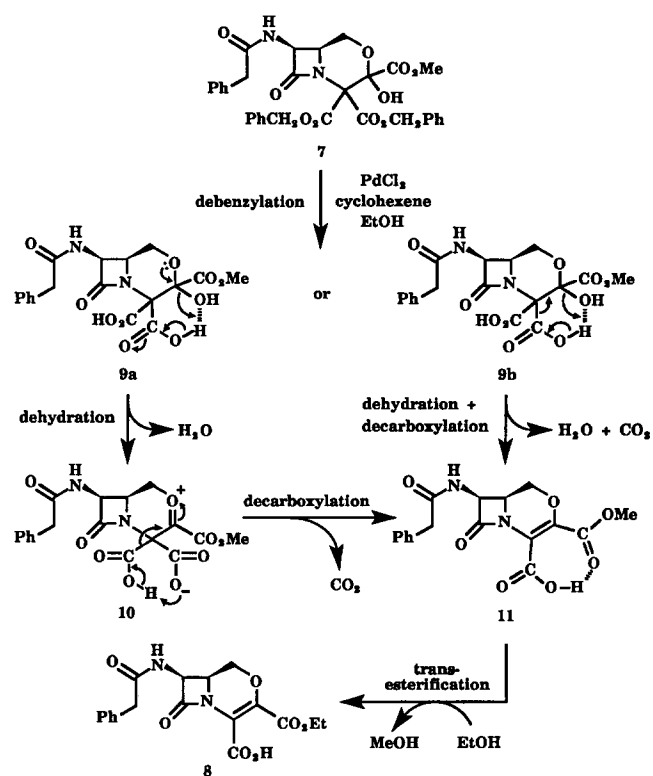


Scheme 1. (a) 95%: *n*-Bu₄NF, THF; (b) 83% overall yield for **2** → **3** → **4** (23% overall yield) + **5** (60% overall yield): (1) 4-methylmorpholine *N*-oxide, tetrapropylammonium perruthenate, CH₂Cl₂; (2) HOCH₂CH₂OH, *p*-TsOH, C₆H₆; (c) 46%: (1) O₃, N₂, CH₂Cl₂; (2) Me₂S; (3) NaBH₄–Al₂O₃; (d) 27% overall yield for **6** → **7** → **8**: (1) 95% aq CF₃CO₂H; (2) PdCl₂, cyclohexene, EtOH.

Key words: β -lactam, isooxacephem, antibiotic, antimicrobial, [1,4]oxazine.

was then treated with Me_2S and NaBH_4 -impregnated alumina⁶ in CH_2Cl_2 . Hydrolysis of acetal **6** with 95% of aqueous $\text{CF}_3\text{CO}_2\text{H}$ afforded lactol **7**, which was subsequently treated with PdCl_2 and cyclohexene in ethanol^{7–10} to give the final product **8** as a racemic mixture in 27% overall yield.

We propose a mechanism for the conversion of lactol **7** to isooxacephem **8** in Scheme 2. Transformation of **7** to **8** by our 'one-flask' method involved debenzylations, dehydration, decarboxylation, and transesterification. After debenzylations occurred in **7**, the internal acid-catalysed dehydration of **9a** took place through a six-membered ring intermediate with anchimeric assistance from the lone pair electrons of the oxygen atom in the 3,4-dihydro-2*H*-[1,4]oxazine ring. Promoted by the oxonium center, the resultant intermediate **10** underwent decarboxylation to give ester **11**. Transesterification of **11** with the solvent, ethanol, produced



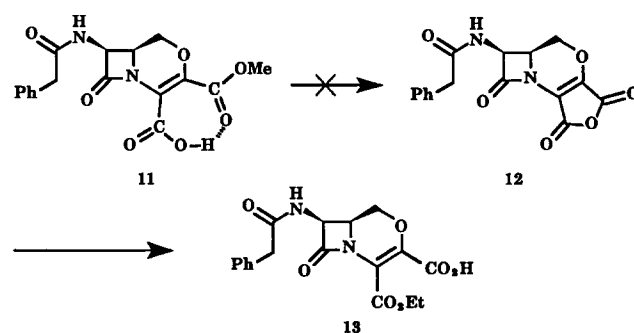
Scheme 2.

the desired product **8**. Alternatively, the conversion of **7** to **8** may be achieved via intermediate **9b** through a concerted mechanism, involving simultaneous dehydration and decarboxylation (Scheme 2). The absence of regioisomeric carboxylic acid **13** in the final product indicates that the alternative route leading **11** to **8** via anhydride **12** did not take place (see Scheme 3).

Biological activity of isooxacephem (**8**)

We tested the biological activity of the synthesized β -lactam (\pm)-**8** as well as naturally occurring penicillin (+)-**14** and reference cephalosporins (+)-**15**,^{11,12} (+)-**16**,¹¹ and (+)-**17**^{13,14} in vitro against several pathogenic microorganisms up to a level as high as 128 $\mu\text{g/mL}$ (Table 1).¹⁵ Our synthetic β -lactam **8** was in racemic form, whereas the comparative compounds (+)-**14**, (+)-**15**, (+)-**16**, and (+)-**17** are single enantiomers.¹⁶ Thus, only one-half of the minimal inhibitory concentration may be necessary for the desired single enantiomer of **8**.

Isooxacephem **8** bearing an ethoxycarbonyl group exhibited notable antimicrobial activity. This might indicate that the electronic activation of the β -lactam moiety by an electron-withdrawing group (e.g., an ester functionality) plays an important role in the biological activity of bicyclic β -lactams.^{17,18} In comparison with the reference penicillin and cephalosporins currently used in medicine, the novel isooxacephem **8** exhibited much stronger activity against methicillin-resistant *S. aureus* 95 and β -lactamase producer *P. aeruginosa* 18S-H (see Table 1).^{19,20}



Scheme 3.

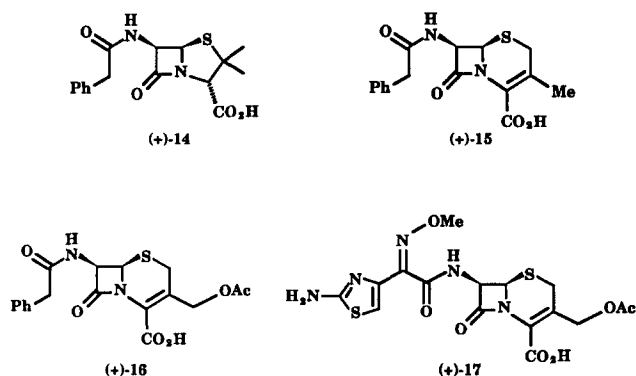
Table 1. Minimal inhibitory concentrations^a ($\mu\text{g/mL}$) of β -lactams (\pm)-**8**, (+)-**14**, (+)-**15**, (+)-**16**, and (+)-**17**

Microorganisms	(\pm)- 8	(+)- 14	(+)- 15	(+)- 16	(+)- 17
<i>S. aureus</i> 95 ^{b,c}	6.92	> 128	> 128	> 128	38.00
<i>S. aureus</i> FDA 209P	0.02	0.40	0.64	0.72	0.08
<i>E. coli</i> ATCC 39188	0.37	2.30	13.13	98.95	0.25
<i>S. typhi</i> O-901	2.50	> 128	24.50	25.32	20.30
<i>P. aeruginosa</i> 18S-H ^c	18.80	> 128	> 128	> 128	128
<i>P. aeruginosa</i> 1101-75	15.00	> 128	100.00	105.30	62.35
<i>K. pneumoniae</i> NCTC 418	0.27	> 128	2.98	15.68	10.25

^a They were obtained by use of a serial broth dilution method and represent the average of triplicate determinations.

^b Methicillin-resistant.

^c β -Lactamase producer.



Experimental

General

Chemicals were purchased from Fluka Chemical Co. Reagent-grade solvents were distilled and then stored over molecular sieves 4 Å. Products were isolated by use of column chromatography (Merck silica gel 60, particle size 230–400 mesh ASTM) packed in glass column (35 g of silica gel/gram of crude material). Analytical TLC analyses were performed on precoated plates (silica gel 60 F₂₅₄) purchased from Merck. Melting points were obtained by a Büchi 510 apparatus. Proton NMR spectra were obtained on a Bruker WH-90 or a Varian XL-200 spectrometer. IR spectra were measured on a Beckman-IR-8 spectrophotometer. Microanalysis data were obtained by use of a Perkin–Elmer 240B microanalyzer.

(±)-Dibenzyl 2-[cis-2-oxo-3-(phenylacetamido)-4-styryl-1-azetidiny]-2-[hydroxy(methoxycarbonyl)methyl]-malonates (diastereomeric mixture; 2). To a solution of **1**² (0.79 g, 1.0 mmol) in THF (35 mL) was added *n*-Bu₄NF (1.0 M solution in THF, 1.0 mL, 1.0 mmol) at 0 °C in portions within 3 min. The solution was stirred at the same temperature for 1 h and was then partitioned between Et₂O (30 mL) and aq KH₂PO₄ buffer soln (pH 4.5, 30 mL). The ethereal layer was dried over MgSO₄ (s) and condensed under reduced pressure. The residue was purified by use of column chromatography (EtOAc as the eluant) to give **2** (0.64 g, 0.95 mmol) as a foam in 95% yield: ¹H NMR (CDCl₃): δ 3.49, 3.51 (2 s, 2H, CH₂CO), 3.65–3.66 (2 s, 3H, CH₃O), 4.21 (br, 1H, OH, exchanged with D₂O), 4.99 (br s, 1H, CH), 5.01–5.30 [m, 5H, 2 × CH₂O + HC(4)], 5.59 [dd, 1H, *J* = 5.0 and 8.5 Hz, HC(3)], 6.22–7.01 (m, 2H, CH=CH), 7.20–7.51 (m, 21H, 4 × C₆H₅ + NH); IR (CH₂Cl₂) *v*_{max}: 3350–3420 (NH, OH), 1778–1745 (β-lactam, ester), 1681 (amide) cm⁻¹. Anal. calcd for C₃₉H₃₆N₂O₉: C, 69.22; H, 5.36; N, 4.14. Found: C, 69.12; H, 5.28; N, 4.21.

(±)-Dibenzyl 2-[cis-2-oxo-3-(phenylacetamido)-4-styryl-1-azetidiny]-2-[ethyleneketal(methoxycarbonyl)methyl]-malonate (4). To a solution of **2** (676 mg, 0.999 mmol) in CH₂Cl₂ (35 mL) containing molecular sieves 4 Å (2.00 g) and 4-methylmorpholine *N*-oxide (175 mg, 1.49 mmol) was added tetrapropylammonium perruthenate (87.7 mg, 0.250 mmol) at room temperature. The

resultant green mixture was stirred for 2 h. Then the reaction mixture was diluted with CH₂Cl₂ (20 mL), washed with 5% aq Na₂SO₃ soln (40 mL), brine (30 mL), and 3% aq CuSO₄ soln (40 mL). The organic layer was dried over MgSO₄ (s), filtered, and condensed under reduced pressure. The residue containing **3** was dissolved in dry benzene (50 mL) containing ethylene glycol (101 mg, 16.2 mmol) and a catalytic amount of *p*-toluenesulfonic acid. The solution was heated at 80 °C for 15 h and the water generated was removed by a Dean–Stark trap. The solution was then cooled, dried over MgSO₄ (s), filtered, and condensed under reduced pressure. After purification by use of a column chromatography (CHCl₃ followed by CHCl₃:EtOAc, 1:1, as the eluant), **4** (165 mg, 0.230 mmol, 23% yield) and **5**⁴ (353 mg, 0.599 mmol, 60% yield) were obtained separately as a foam.

For **4**: ¹H NMR (CDCl₃): δ 3.55 (s, 2H, CH₂CO), 3.71–4.18 (m, 7H, CH₃ + OCH₂CH₂O), 5.00–5.29 [m, 5H, 2 × CH₂O + HC(4)], 5.52 [dd, 1H, *J* = 5.0 and 9.0 Hz, HC(3)], 6.12–6.95 (m, 3H, CH=CH + NH), 7.02–7.46 (m, 20H, 4 × C₆H₅); IR (CH₂Cl₂) *v*_{max}: 3405 (NH), 1779–1750 (β-lactam, ester), 1680 (amide) cm⁻¹. Anal. calcd for C₄₁H₃₈N₂O₁₀: C, 68.51; H, 5.33; N, 3.90. Found: C, 68.40; H, 5.31; N, 4.01.

(±)-Dibenzyl 2-[cis-4-hydroxymethyl-2-oxo-3-(phenylacetamido)-1-azetidiny]-2-[ethyleneketal(methoxycarbonyl)methyl]malonate (6). β-Lactam **4** (718 mg, 0.999 mmol) in CH₂Cl₂ (70 mL) was saturated with N₂ gas at –78 °C. Then a mixed O₃ and N₂ gas was bubbled in until KI starch paper turned dark blue, which indicated the presence of excess O₃. The excess O₃ was removed by a stream of N₂ gas. Then Me₂S (620 mg, 9.98 mmol) was added at –30 °C. After 1 h, NaBH₄:Al₂O₃ (1:10 g/g) was added and the reaction mixture was stirred at room temperature for 1 h. Then it was filtered and condensed under reduced pressure. After purification by use of a column chromatography (EtOAc as the eluant), **6** (297 mg, 0.460 mmol) was obtained as a foam in 46% yield: ¹H NMR (CDCl₃): δ 3.50 (s, 2H, CH₂CO), 3.70–5.40 (m, 15H, CH₃ + OCH₂CH₂O + CHCH₂OH + 2 × CH₂O), 5.58 [dd, 1H, *J* = 5.0 and 9.0 Hz, HC(3)], 6.92 (br, 1H, NH), 7.00–7.42 (m, 15H, 3 × C₆H₅); IR (CH₂Cl₂) *v*_{max}: 3300–3410 (NH, OH), 1778–1740 (β-lactam, ester), 1680 (amide) cm⁻¹. Anal. calcd for C₃₄H₃₄N₂O₁₁: C, 63.15; H, 5.30; N, 4.33. Found: C, 63.20; H, 5.29; N, 4.48.

(±)-3-Ethyl 2-hydrogen (6*RS*,7*RS*)-8-oxo-7-(phenylacetamido)-4-oxa-1-azabicyclo[4.2.0]oct-2-ene-2,3-dicarboxylate (8). Hydroxyl acetal **6** (2.58 g, 3.99 mmol) was dissolved in 95% aq CF₃CO₂H (50 mL) and the solution was stirred at room temperature for 7 h. It was then poured into a mixture of crushed ice and brine, and then was extracted with EtOAc. After the organic solution was washed with water (2 × 40 mL) and dried over MgSO₄ (s), the solvent was removed to afford **7** as an oil. The crude **7** was then dissolved in EtOH (60 mL), to which were added PdCl₂ (400 mg,

2.26 mmol) and cyclohexene (45 mL). The mixture was heated at reflux for 6 h, cooled, filtered, and the residue was washed with 80% aq EtOH (2 × 20 mL). The combined filtrates and washings were condensed to dryness under reduced pressure. After purification by use of column chromatography (EtOAc:MeOH, 9:1, as the eluant), **8** (403 mg, 1.08 mmol) was obtained in 27% yield: mp 138–140 °C (from ether); ¹H NMR (DMSO-*d*₆/CDCl₃/D₂O): δ 1.34 (t, 3H, *J* = 7.0 Hz, CH₃), 3.50 (s, 2H, CH₂CO), 3.45–3.89 [m, 2H, H₂C(5)], 4.20 (q, 2H, *J* = 7.0 Hz, CH₂O), 4.42 [m, 1H, HC(6)], 5.01 (d, 1H, *J* = 5.0 Hz, CHND), 7.40 (s, 5H, C₆H₅); IR (CH₂Cl₂) ν_{max}: 3050–3652 (COOH, NH), 1792 (β-lactam), 1748 (ester), 1715, 1708 (acid, C=C), 1680 (amide) cm⁻¹. Anal. calcd for C₁₈H₁₈N₂O₇: C, 57.75; H, 4.85; N, 7.48. Found: C, 57.68; H, 4.98; N, 7.34.

Antimicrobial activity tests

The serial broth dilution method was used to study the antibiotic activity.¹⁵ The inocula were prepared by use of the heart infusion broth (Difco Laboratories) to make 10⁻⁴ dilutions of the overnight cultures. Tubes of the seeded antibiotic-containing media were incubated at 37 °C for 20 h. The lowest concentration of antibiotic that prevented visible growth of microorganisms was then determined.

Acknowledgments

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